Validation of extraction methods for total RNA and miRNA from bovine blood prior to quantitative gene expression analyses Andrea Hammerle-Fickinger, Irmgard Riedmaier, Christiane Becker, Heinrich H. D. Meyer, Michael W. Pfaffl, Susanne E. Ulbrich Physiology Weihenstephan, Technische Universitaet Muenchen, Weihenstephaner Berg 3, 85354 Freising, Germany Corresponding author: Susanne E. Ulbrich Physiology Weihenstephan, Technische Universitaet Muenchen Weihenstephaner Berg 3, 85354 Freising Germany Phone: +49 8161 71 4429 Fax: +49 8161 71 4204 ulbrich@wzw.tum.de Journal: Biotechnology Letters Journal section: Animal Cell Technology

Supplementary Data

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- 32 Extraction of total RNA from whole blood after blood collection (WB) and from
- 33 <u>leukocytes after lysis of erythrocytes (LY)</u>
- 34 Total RNA was extracted from samples homogenized in TriFast with minor
- 35 modifications according to the manufacturer's instructions (Peqlab Biotechnologie).
- 36 Therefore samples were kept for 30 min at room temperature until complete thawing.
- 37 For phase separation 200 μl chloroform (-20 °C) for 1 mL TriFast (originally used) was
- added following vortexing for 10 sec, and incubation at room temperature for 10 min.
- 39 Centrifugation was done at 12000 rpm for 15 min at 4 °C. RNA containing phase was
- 40 transferred to a fresh tube and RNA precipitation was performed adding 500ul
- 41 isopropanol (-20°C) for 500µl of RNA aqueous phase. Samples were mixed,
- 42 incubated at room temperature for 10 min and centrifuged for 10 min at 4 °C at
- 43 12000 rpm. Supernatant was carefully removed and wash of the RNA pellet with
- 44 500µl of 75% ethanol by vortexing and subsequent centrifugation for 5 min at 4 ℃ at
- 45 12000 rpm was performed twice. To remove excess of alcohol, samples were placed
- 46 in a heat block at 50 ℃. Solubilisation of RNA pellet was done using Rnase-free
- 47 water pipetting it up and down following 10 min of heating at 50 °C and storage at 80
- 48 °C.
- 49 Extraction of mRNA and miRNA from leukocytes stabilized by LeukoLOCK (LL)
- 50 Locked cells in filter cartidges were flushed with 2.5 ml of mirVana miRNA Lysis
- 51 Solution and collected in 15 ml conical tubes. To this solution 1/10 miRNA
- 52 Homogenate Additive was added and vortexed. Filter cartidges were flushed with 1.5
- 53 ml acid phenol and collected in the same tube. Solution was vigorously vortexed for

54 30 sec, kept 5 min at room temperature and centrifuged for 10 min at 3200 rpm. 55 Supernatant was carefully removed and transferred to a fresh 15 ml tube, mixed with 1.25 volumes of ethanol. Solution was multiple loaded over filter cartidge, centrifuged 56 57 at room temperature for 15 sec at 10000 rpm and flow-through was collected in a fresh 15 ml tube and saved at 4 °C for miRNA recovery. To the filter 700 μl of miRNA 58 59 Wash Solution was applied and centrifuged at room temperature for 10 sec at 10000 rpm. Flow-through was discarded and filter was washed twice with 500µl Wash 60 61 Solution following centrifugation at room temperature for 1 minute at 10000 rpm. RNA was recovered applying 100 µL of pre-heated (95°C) Elution Solution to the 62 63 filter, centrifuged for 30 sec at 12000 rpm and storage at -80 ℃. To the Saved flow-through containing the miRNA portion 2/3 volume of 100% ethanol 64 was added and mixed thoroughly. Solution was multiple loaded over a filter cartridge, 65 66 centrifuged at room temperature for 15 sec at 10000 rpm and flow-through was discarded. To the filter 700 µl of miRNA Wash Solution was applied and centrifuged 67 68 at room temperature for 10 sec at 10000 rpm. Flow-through was discarded and filter 69 was washed twice with 500µl Wash Solution following centrifugation at room temperature for 1 minute at 10000 rpm. miRNA was recovered applying 100 µL of 70 71 pre-heated (95°C) Elution Solution to the filter, centrifuged for 30 sec at 12000 rpm 72 and storage at -80 °C.

Extraction of mRNA and miRNA from leukocytes after blood fractionation (PI)

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- 74 Total RNA was extracted from samples homogenized in Qiazol reagent (Qiagen).
- 75 Therefore samples were kept for 30 min at room temperature until complete thawing.
- 76 For phase separation, 140 µl chloroform was added following vortexing for 15 sec
- and incubation at room temperature for 3 min. Centrifugation was done at 12000 rpm
- 78 for 15 min at 4 °C. RNA containing phase was carefully transferred to a fresh tube
- and RNA precipitation was performed adding 1.0 volume of 70% ethanol. Solution

was multiple loaded over RNeasy Mini Spin filter, centrifuged at room temperature for 15 sec at 10000 rpm and flow-through was collected in a fresh 15 ml tube and saved at 4 ℃ for miRNA recovery. To the RNeasy Mini Spin filter 700 µl Buffer Wash Solution was applied and centrifuged at room temperature for 15 sec at 10000 rpm. Flow-through was discarded and filter was washed twice with 500µl Buffer Solution following centrifugation at room temperature for 15 sec at 10000 rpm. RNA was recovered applying 30 µL of RNA free water to the filter, centrifuged for 1 minute at 10000 rpm and storage at -80 °C. To the Saved flow-through containing the miRNA portion 2/3 volume of 100% ethanol was added and mixed thoroughly. Solution was multiple loaded over a MinELute filter cartridge, centrifuged at room temperature for 15 sec at 10000 rpm and flow-through was discarded. Filter was washed with 500µl Buffer Solution following centrifugation at room temperature for 15 sec at 10000 rpm. Following wash step was done applying 500 µl of 80% ethanol to the filter following centrifugation at room temperature for 2 min at 10000 rpm. miRNA was recovered applying 14 µL of RNA free water to the filter, centrifuged for 1 minute at 10000 rpm and storage at -80 °C. Extraction of mRNA and miRNA from whole blood collected in PAXgene tubes (PAX Samples were thawed in a wire rack at ambient temperature for 2 hours. After PAXgene tube centrifugation for 10 min at 3000 rpm the supernatant was discarded. To the pellet 5 ml RNA free water was added, vortexed, centrifuged for 10 min at 3000 rpm and supernatant was discarded. For dissolving the pellet 360 µl of Buffer BR1 was used and solution was transferred into a 1.5 ml tube. To this solution 300 µl Buffer BR2 and 40 µl Proteinase K was added, vortexed and incubated for 10 min at 55 °C using a shaker incubator at 1400 rpm. Samples were centrifuged at 13000 rpm and supernatant was transferred to a fresh tube. Precipitation was done adding 350 μl of 100% ethanol, mixing and centrifuged briefly. Solution was multiple loaded over

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a PAXgene spin column, centrifuged at room temperature for 1 min at 10000 rpm and flow-through was collected in a fresh 15 ml tube and saved at 4 °C for miRNA recovery. To the PAXgene spin column filter 700 μl Buffer BR3 was applied and centrifuged at room temperature for 1 min at 10000 rpm. Flow-through was discarded and filter was washed twice with 500μl Buffer BR4 following centrifugation at room temperature for 3 min at 13000 rpm. RNA was recovered applying 40 μL of Buffer BR5 directly onto the filter, centrifuged for 1 minute at 10000 rpm and storage at -80 °C.

To the Saved flow-through containing the miRNA portion 1.4 volume of 98% ethanol was added and mixed thoroughly. Solution was multiple loaded over a RNeasy mini spin column, centrifuged at room temperature for 1 min at 13000 rpm and flow-through was discarded. Filter was washed twice with 500μl Buffer Solution following centrifugation at room temperature for 15 sec at 13000 rpm. miRNA was recovered applying 30 μL of RNA free water to the filter, centrifuged for 1 minute at 13000 rpm

and storage at -80 °C.